

BIOSYNTHESIS OF OXANTHROMICIN

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Oxanthromicin (Antibiotic 16-550), a novel dimeric anthrone peroxide produced by *Actinomadura* sp. SCC 1646, exhibited good *in vitro* activity against dermatophytic fungi. The details of taxonomy, biological properties¹⁾ and the structure²⁾ have been reported from our laboratories. The biosynthetic origin of aclacinomycins³⁾, cetocycline⁴⁾, tetracyclines⁵⁾ and of similar ring structures has been studied extensively and their overall polyketide mechanism is well established⁶⁾. However oxanthromicin is a unique structure and we report here our studies on the biosynthesis of the basic nucleus.

Addition of ¹⁴C-labeled Precursors

The fermentation was carried out either in flasks or in a 14-liter New Brunswick Scientific Laboratory fermentor according to the published procedure¹⁾.

The optimum addition time of the labeled precursors, was studied using L-[methyl-¹⁴C]-methionine, Na-[1-¹⁴C]acetate, Na-[2-¹⁴C]acetate, and Na-[1-¹⁴C]propionate. The additions (conc 2.5 μ Ci/10 ml) were made at 24, 48 and 72 hours of fermentation. Fermentations were harvested at 96 hours, pH adjusted to 5, and extracted twice with EtOAc. The extract was concentrated 20-fold under vacuum. The amount of radioactivity incorporated was measured in a scintillation spectrophotometer. The EtOAc extract (100 μ l) was chromatographed on TLC silica plates (Rf 0.33) in CHCl₃ - MeOH - 17% ammonia (2:2:1) and then scanned for radioactivity on a Nuclear Chicago radiochromatogram scanner (Model 1002).

The rate of incorporation of the precursors

Table 1. Incorporation of ¹³C-labeled precursors into oxanthromicin.

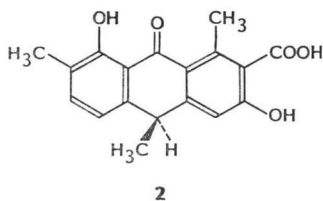
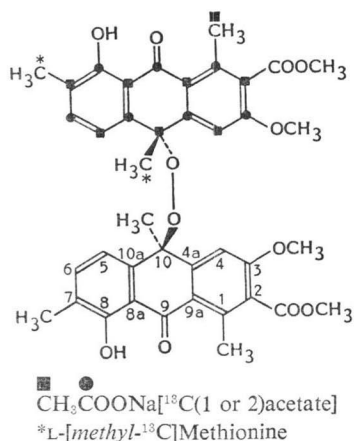
Carbon atoms	Chemical shift ^b ppm (δ)	Enrichment factor ^a		
		[1- ¹³ C]- Acetate	[2- ¹³ C]- Acetate ^c	L-[methyl- ¹³ C]- Methionine
1	141.3	1.69	1.13	0.97
2	127.0	1.20	1.51	1.12
3	159.1	2.39	1.36	1.05
4	117.5	0.76	0.95	1.28
4a	150.9	1.71	1.20	0.93
5	107.3	0.73	0.98	1.31
6	135.9	1.61	0.85	1.11
7	115.3	1.11	1.32	1.15
8	160.8	2.05	1.03	0.92
8a	122.4	1.20	1.21	1.19
9	189.6	1.67	1.13	0.93
9a	125.9	0.86	1.21	1.07
10	79.8	0.94	1.33	1.23
10a	140.6	2.13	1.22	1.10
1-CH ₃	20.8	0.85	1.17	1.09
2-COOCH ₃	168.1	2.31	1.20	0.86
2-COOCH ₃	56.1	Reference	—	—
3-OCH ₃	52.5	Reference	—	—
10-CH ₃	34.0	0.85	1.10	1.10
7-CH ₃	15.6	0.79	1.14	1.03
				14.8
				14.6

^a Peak heights of the enriched sample divided by the natural abundance sample obtained from spectra run under essentially identical instrumental conditions. The average of peak intensities due to OCH₃ carbons, is taken as 1.00 and the heights of other peaks are relative to 1.00.

^b The chemical shift assignments are based upon single frequency off-resonance (SFOR) and single frequency proton-decoupled spectra.

^c Results of two separate experiments.

Fig. 1. Biosynthesis of oxanthromicin.



The addition of $[1-^{13}\text{C}]$ acetate enriched 8 carbons; C-1, C-3, C-4a, C-6, C-8, C-9, C-10a and C(2)- COOCH_3 . Similarly the addition of $[2-^{13}\text{C}]$ acetate* enriched 8 carbons; C-2, C-4, C-5, C-7, C-8a, C-9a, C-10 and C(1)- CH_3 , whereas, L- $[methyl-^{13}\text{C}]$ methionine only enriched two carbons; C(10)- CH_3 and C(7)- CH_3 .

The results of the incorporation experiments are consistent with a biosynthetic pathway involving polyketide formation, alkylation, reduction and dehydration to give the monomer **2**, which is then oxidatively converted into the natural product (Fig. 1). The nature of this final conversion, which is presumably an enzyme-mediated transformation as the product is optically active, is of moderate interest.

Direct evidence for the intermediacy of **2** will require further studies with labeled substrate.

Addition of ^{13}C -Labeled Substrate

At the optimum times L- $[methyl-^{13}\text{C}]$ methionine, Na- $[1-^{13}\text{C}]$ acetate, and Na- $[2-^{13}\text{C}]$ acetate were added individually (conc 250 mg/liter) to the culture. The fermentation broth was harvested at 96 hours, acidified to pH 5 with conc HCl, and extracted twice with EtOAc. The organic extract was concentrated to about 1 liter, washed with 0.1 N HCl, several times with H_2O , dried over MgSO_4 and finally evaporated to dryness. The dried material was washed with hexane to yield crude antibiotic.

The crude antibiotic was methylated with CH_2N_2 in ether for 4 hours and the excess CH_2N_2 decomposed with dil AcOH. The ether phase was separated, washed with H_2O , dried over MgSO_4 , evaporated, and the solid residue purified by preparative TLC, on silica plates.

Proton-noise decoupled (p.n.d.) ^{13}C NMR spectra of the natural abundance ^{13}C and the enriched tetramethylated antibiotic samples were recorded in CDCl_3 on a Varian XL-100-15 NMR spectrometer operating at 25.2 MHz. Percentage enrichments are shown in Table 1.

The ^{14}C -labeled studies indicated that only $[1-^{14}\text{C}]$ acetate, $[2-^{14}\text{C}]$ acetate and L- $[methyl-^{14}\text{C}]$ methionine were incorporated into the antibiotic.

References

- 1) PATEL, M.; A. C. HORAN, V. P. GULLO, D. LOEBENBERG, J. A. MARQUEZ, G. H. MILLER & J. A. WAITZ: Oxanthromicin, a novel antibiotic from *Actinomadura*. *J. Antibiotics* 37: 413~415, 1984
- 2) WRIGHT, J. J.; Y. MERRILL, M. S. PUAR & A. T. MCPHAIL: Structure of oxanthromicin (antibiotic 16-550), a novel dimeric anthrone peroxide. *J. Chem. Soc. Chem. Commun.* 1984: 473~474, 1984
- 3) KITAMURA, I.; H. TOBE, A. YOSHIMOTO, T. OKI, H. NAGANAWA, T. TAKEUCHI & H. UMEZAWA: Biosynthesis of aklavinone and aclacinomycins. *J. Antibiotics* 34: 1498~1500, 1981
- 4) MITSCHER, L. A.; J. K. SWAYZE, T. HÖBERG, I. KHANNA, G. S. RAGHAV RAO, R. J. THERIAULT, W. KOHL, C. HANSON & R. EGAN: Biosynthesis of cetocycline. *J. Antibiotics* 36: 1405~1407, 1983
- 5) THOMAS, R. & D. J. WILLIAMS: Oxytetracycline biosynthesis. Mode of incorporation of $[1-^{18}\text{C}, ^2\text{H}_3]$ acetate. *J. Chem. Soc. Chem. Commun.*

* The reasons for the low incorporation of $[2-^{13}\text{C}]$ acetate (Table 1) are not fully understood. However, the results of two experiments support the enrichment of 8 carbons.

- 1984: 443~444 and earlier references, *ibid*,
1983: 128~130 and 677~679, 1983
- 6) SIMPSON, T. J.: The biosynthesis of polyketides.
Natural Product Reports 1984: 281~297, 1984